The Antimicrobial Activity of Quinolones

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Abstract- The creation of drugs with increased action against Gram-positive bacteria has been a current focus of quinolone antibacterials. A greater understanding of the structural characteristics of quinolones that provide the optimal mix of clinical performance and decreased resistance selection in Gram-positive bacteria has been made possible by considerable research efforts. With an understanding of their chemical structure and how this affects target specificity, avoiding efflux, and preventing the emergence of quinoloneresistant mutants in Gram-positive bacteria, this review takes into account the structural features of new quinolones and their relationship to antibacterial activity against Grampositivebacteria.

Keywords- Quinolone, fluoroquinolone, gram-positive bacteria, gram, negative bacteria, antimicrobial activity.

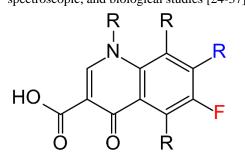
I. INTRODUCTION

The rise of gram-positive bacteria that are multidrug resistant, such as methicillin-resistant Staphylococcus aureus (MRSA), has made treating infectious diseases challenging and has, over the past few decades, developed into a significant medical issue.

The greatest strategy to combat bacterial resistance and provide efficient therapies is to find novel, potent antibacterial medications because pathogenic bacteria are constantly evolving mechanisms of resistance to the antibacterials that are currently being utilized [1].

Enhancing the pharmacokinetic characteristics and raising the activity against gram-positive cocci and anaerobes have been key quinolone research strategies in recent years [2-5].Quinolones may result in negative side effects including hypoglycemia, severe heart dysrhythmias, and tendinitis, phototoxicity, and CNS effects [6, 7].The most frequent substitution at position C-7 is a five- or six-membered ring; examples include theaminopyrrolidinesubstituent found in gemifloxacin and trovafloxacin [8, 9].Theantibacterial efficacy of several fluoroquinolones with an oxime or a substituted oximeconnected to the pyrrolidine or piperazine ring at C-7

position was also Investigated [10-15]. The efficient suppression of DNA replication is primarily responsible for quinolones' antibacterial action. The theorized process for how quinolones interact with metal cation was described as chelation between the metals, the 4-oxo group, nearby carboxyl groups [16]. is crucially important to design new drugs that are effective against resistant species. The new generation of quinolones antibacterial agents significantly improved in terms of potency, spectrum, and pharmacokinetic properties, however these agents had to contend with a sharp rise in resistance from Gram-positive pathogens [17, 18]. It is noteworthy that alkyl groups including ethyl, vinyl, cyclopropyl, and tert-butyl groups have been considered ideal N-1 substituents since they play a significant role in the fluoroquinolonesantibacterial action. It is also known that the fluoroquinolones' bulkiness and the stereochemistry of their N-1 substituent play a key role in the antibacterial activity of these compounds [19].Additionally, it is used to treat prostatitis, skin and tissue infections, urethral and cervical gonococcal infections, pyelonephritis, sexually transmitted diseases, urinary tract infections, and pyelonephritis [20-23]. In an effort to better understand the binding mode and effects. potential synergistic numerous transitionmetalcomplexes with different quinolone antibacterial medicines have undergone extensive structural, spectroscopic, and biological studies [24-37].



II. ANTIMICROBIAL ACTIVITY

• Agar-dilution testing was used to assess the antibacterial activity of various compounds. The compounds and reference medications were produced

in Mueller Hinton agar using twice-serial dilutions. Drugs were dissolved in DMSO, then water was added to dilute the solution. Colonies from Mueller-Hinton agar media that had grown overnight were suspended in saline to create the bacterium inoculate. To get 107 CFU/mL, the samples were then diluted in 0.85% saline. Spot-inoculating petri dishes with 1 IL of each prepared bacterial suspension (104 CFU/spot) allowed for an 18-hour incubation period at 35–378°C. There was no discernible growth on the plate because the MIC was the lowest concentration of the test substance. Using test media as the control, a test was conducted to make sure the solvent had no impact on bacterial growth [38].

- newly synthesized The N-(5-benzylthio-1,3,4thiadiazol-2-yl) N-(5-benzylsulfonyl-1,3,4and thiadiazol-2-yl)piperazinyl quinolone derivatives were screened with Gram-positive (Staphylococcus aureus ATCC 6538p, Staphylococcus epidermidis ATCC 12228, and Bacillus subtilis PTCC 1023) and Gram-negative (Escherichia coli ATCC 8739, Klebsiellapneumoniae ATCC 10031. and Pseudomonas aeruginosa ATCC 9027) bacteria were tested against compounds in vitro using the traditional agar- In order to calculate the minimum inhibitory concentrations (MIC), parent quinolones, norfloxacin, and ciprofloxacin were used as benchmarks. According to the MIC values, the majority of the substances had significant activity (MIC = 0.03-4 lg/ mL) against Gram-positive bacteria and moderate to poor activity (MIC = 1-64lg/mL) against Gram-negative pathogens [39].
- Each test substance was dissolved in a suitable solvent (100 percent DMSO), then serially diluted with DMSO to the required testing concentration ranges. The 48-well plate with 0.99 mL of medium broth containing 1-5 105 CFU/mL testing microorganism received each series of testing solution (0.01 mL). Thus the final maximal concentration of\sDMSO was 1%, and the initial concentration of testing solution\swas 300 iM. The following media were used: nutrient broth (NB, for Escherichia coli, Pseudomonas DIFCO) aeruginosa, and Klebsiellapneumoniae; Mueller-Hinton broth (DIFCO) for S. aureus, methicillinresistant (MRSA), and Proteus vulgaris; brain heart infusion broth (BHI, DIFCO) for Mycobacterium ranae; and tryptic soy broth. The plates were incubated for 20-72 hours at 37 °C, and the MIC was

calculated either visually by measuring turbidity or indirectly by observing microbial growth under a microscope. Every test employed vehicle and reference compounds as the negative and positive controls, and the assays were run twice.

- Staphylococcus aureus, Escherichia coli, and Pseudomonas aeruginosa were used as test organisms to determine the antibacterial activity of the compounds (ligand and complexes) (P. aeruginosa). The microorganisms were kindly provided by the Department of Medical Laboratories, Technological and Educational Institution of Larissa, Greece, and were kept at the laboratory of "Chemical Biology" of NCSR "Demokritos." Dr. I. Eskioglou.
- By figuring out the MIC, screening was carried out. There were two types of media used: the Luria broth medium (=LB, including 1% w/v tryptone, 0.5% w/v NaCl, and 0.5% w/v yeast extract) and the minimal medium salts broth (=MMS, containing 1.5% w/v glucose, 0.5% w/v NH4Cl, 0.5% w/v K2HPO4, 0.1% w/v NaCl, 0.01% w/v M In distilled water, the compounds were dissolved using two-fold serial dilutions ranging from 8 to 0.125 lg mL1. At 37 C, all cultures were incubated. Additionally, control tests were run without any active components.
- Using twofold serial dilutions in liquid media containing 0.125–8 lg mL1 of the substance under test, the MIC was calculated. At the right temperature for each species, a preculture of bacteria was cultivated in LB for the entire overnight period. 20 mL of this preculture was infected with two mL of MMS. To determine whether the growth of the bacterium under test is normal, this culture was utilised as a control. A similar second culture was created by adding 20 IL of the bacteria together with the tested substance at the desired concentration.
- To test the substance's impact on MMS, a second control sample was employed, consisting of 2 mLMMS added with the same compound concentration. Every sample was measured twice. By monitoring the turbidity of the culture after 12 and 24 hours, we were able to track bacterial growth. Half of a compound's concentration was examined to see if it could stop bacterial growth at a specific concentration. This process was continued until the concentration at which bacteria ordinarily grow. The MIC value was established as the lowest concentration at which bacterial growth was inhibited. The tools and culture media were all clean.

- When tested against two Gram (-), E. coli and P. aeruginosa, and one Gram (+), S. aureus, microorganisms, the efficacy of the ligand and the complexes was determined. The findings are shown in the ligand and complexes, which exhibit inhibitory activity against all of the tested microorganisms. Additionally, the coordination of enrofloxacin (MIC = 1-8 lg mL1) with transition metal ions results in increased biological activity (MIC for the complexes = 0.25-4 lg mL1). Investigations have also been done on the metal salts' antibacterial properties.
- The concentration range used to test the activity of the complexes in this work revealed that the metal salts do not display antibacterial activity. The presence of the enrofloxacinato ligands in the molecule is primarily responsible for the complexes' low MIC values. The following essential considerations should be made while examining the antibacterial activity of metal complexes: I the ligands' chelate action; (ii) the N-donor ligands' makeup; (iii) the complex's overall charge; (iv) the presence and make-up of the ion that neutralises the ionic complex; and (v) the nuclearity of the metal centre in the complex. The chelate effect of the ligand is only apparent for complexes as the first component [40-43]. This is most likely one of the causes of the complexes' varied antibacterial activity, though this diversity may also be significantly influenced by the nature of the metal ion coordinated to the enrofloxacinato ligand.
- In general, all the complexes exhibit better inhibition\sthan free enrofloxacin against S. aureus and equal or better inhibition against E. coli and P. aeruginosa. In this study, Fe (erx)3 exhibits the best inhibition of all the complexes and is four times more effective than enrofloxacin against all of the microorganisms used, indicating that the coordination of the enrofloxacinato ligand to Fe3+ has increased its antimicrobial activity [44, 45]. However, the remaining complexes are two to four times more effective than enrofloxacin against S. aureus while they are equally effective against the two Gram(-) bacteria. However, Cu (erx) 2(H2O) (MIC = 0.125 lg mL1) gave the best suppression among all enrofloxacinato complexes identified against the two Gram (-) bacteria.
- The Z- and E-1-(2-fluorovinyl)quinolone derivatives (15 and 16)'s in vitro antibacterial efficacy against two Gram-positive strains lists the results against

two Gram-negative strains (Escherichia coli NIHJ JC-2 and Pseudomonas aeruginosa IID 1210), two Staphylococcus aureus strains (Staphylococcus aureus Smith and Streptococcus pneumoniae type III), and those against three Gram-positive strains.

- In vitro antibacterial activity against both Grampositive and Gram-negative microorganisms was 2to 32-fold stronger for the Z-isomers Z-15a-c and Z-16a-c than it was for the comparable E-isomers. Our are consistent with those of 1-(2results fluorocyclopropyl)quinolone derivatives, whose cis isomers shown stronger antibacterial activity than the trans isomers [46]. Additionally, Z-15a-c and Z-16a-c showed in vitro antibacterial activity that was comparable to that of [46]. The most effective in vitro antibacterial activity of all the compounds tested was that of the 7-(3-aminopyrrolidinyl) derivative Z-16b, which has a fluorine atom at the C-8 position. Z-16b also demonstrated comparable in vitro antibacterial activity against S. aureus Smith, St. pneumoniae type III, and P. aeruginosa IID1210 to that of 2.except for its effectiveness against E. coli NIHJ JC-2.
- Next, we assessed Z-15b's ability to suppress growth.and E-15b, which are analogues of that are conformationally constrained, as well as the inhibitory potentials of 2 and 5 when S. aureus DNA gyrase is present. According to Table 2, Z-15b had an IC50 value that was similar to 5 and nine times smaller than E-15b's. The different in vitro antibacterial activities of Z-15b, E-15b, and 5 against S. aureus Smith were mirrored in the disparities in their inhibitory capacities. The position of the fluorine atom in the 2-fluorovinyl group of Z-15a and E-15b appeared to have an impact on the stereochemistry of the 2-fluorovinyl group.both in vitro antibacterial efficacy and DNA gyrase inhibition as the target enzyme.Using the traditional agar-dilution method, compounds 5a-l had their antibacterial activity assessed [47].
- The substances and reference medications were diluted twice serially inThomson-Muller agar. DMSO (1 ml) was used to dissolve in it.
- The drugs (10.0 mg), and water was added to dilute the solution (9 ml). The necessary concentrations of 100, 50, 25, 12.5, 6.25, 3.13, 1.56, 0.78, 0.39, 0.19, 0.098, 0.049, 0.025, 0.013, 0.006 and 0.003m g/ml were obtained by doing another incremental twofold dilution with melting Mueller-Hinton agar. The bacteria inocula were made by suspending Mueller-

Hinton agar media overnight colonies in 0.85% saline. At 600 nm, the inocula were photometricallyadjusted to a cell density of around 0.5 McFarland standard (1.5108 CFU/ml).To get 107 CFU/ml, the suspensions were then diluted in 0.85% saline. One millilitre of each produced bacterial suspension (104 CFU/spot) was spot-inoculated into petri dishes, which were then incubated at 35–37 °C for 18 hours. the least.

- The test compound's inhibitory concentration (MIC), which was the lowest concentration, prevented any growth from being seen on the plate. A control test was conducted using test medium supplemented with DMSO at the same dilutions as used in the experiment to make sure the solvent had no impact on bacterial growth.
- Using Muller-Hinton agar and the agar dilution method14, the MIC (g/mL) was calculated (Difco Laboratories, Detroit, MI). MIC was specified.as the lowest dose of an antibacterial drug that during an 18-hour incubation period at 35 °C, prevented discernible growth.
- When used against both Gram-positive and Gramnegative bacteria, 7-thiazolyl-4-quinolones showed the strongest biological activity.Unlike the outcomes we saw with the substituted oxazole compounds, activity was observed when the thiazole moiety had aminomethyl substituents.
- This family of chemicals lacked any effective broadspectrum antibacterial action. The Staphylococcus organisms responded best to the most active chemicals [48-51]. In general, the chemicals in this study were more effective against Gram-positive than Gram-negative organisms. In this series, substituting a carbon atom for the nitrogen atom in the quinolone ring system at position seven did not result in quinolone derivatives with strong antibacterial activity. Culbertson et al.
- In medicine, antibacterial compounds from the aforementioned class have been employed. When treating infections of the urinary tract, respiratory tract, and skin, derivatives of 4-oxo-1, 4dihydroquinoline-3carboxylic acids are utilised.intestinal infections, ear, nose, and throat infections, STDs, soft tissue and skin infections, gram-negative and Staphylococcus meningitis, liver and bile infections, septicemia and endocarditis, prophylactic and postoperative infections, and infections in individuals with immune deficiencies.Good antibacterial activity have been

documented against a number of species, including Moraxella, Enterococcus faecalis, and Staphylococcus aureus

- There is no substance being developed from this class molecules, which includes bacteria of like Catarrhalis, Escherichia coli, Haemophilusinfluenzae, and Streptococcus pneumoniae. Interestingly, despite several of them showing a very powerful antibacterial profile in preclinical testing, none of the quinolones with C-7 linkage through C-atom have been successful in reaching the market due to various types of toxicity. [52]. As a result, we thought that by adding a tetrahydrothieno[3,2-c]pyridine moiety through the N-atom of the substituent at the C-7 position, compounds 7-20 would be created that are safer and more effective antibacterial compounds. We previously reported on oxazolidinones that contained tetrahydrothieno[3,2-c]pyridine, and those compounds shown strong antibacterial activity in MIC assays. [53] We report the synthesis, characterization, and8 results in this communication.
- The new fluoroquinolones were evaluated against 26 representative Gram-positive and Gram-negative pathogens for their antibacterial activity. The transmethyl substituted analogue 1 2 and ofloxacin, which served as positive controls, were used as the minimum inhibitory concentrations (MICs) for the produced compounds (OFLX) and a list of tosufloxacin (TFLX) is provided in [54]. The 3-hydroxymethyl-4-trifluoromethyl derivatives (11 a and 11 d) had stronger antibacterial effects against quinolone-resistant SA strains but were less effective against Gram-negative pathogens.
- The 3-amino-4-trifluoromethyl 3.3and its disubstituted derivatives (11b, 1le, and 1lg-i) shown a striking reduction in efficacy against quinoloneresistant SA among the investigated drugs. The 3aminomethyl-4-trifluoromethyl derivatives (11 c and 11 f) had very strong antibacterial action against Gram-positive bacteria, including SA that was quinolone-resistant.Quinolones are used as broad spectrum antibacterial medicines that work against both Gram-positive and Gram-negative bacteria all over the world. Numerous quinolone compounds have antiviral, antimalarial, and anticancer properties. Quinolones developed into a distinct class of synthetic pharmaceuticals after being initially created as a result of chloroquine manufacturing. Different synthetic techniques are now being investigated to create novel quinolone analogues that are more

potent. Some bacteria develop resistance to them as a result of the abuse of these medications, and these bacterial strains are now a common threat globally. There are many explanations for resistance, but a typical one is mutation at the target location. Exploring new synthesis pathways for novel quinolone analogues that are economically viable, less cytotoxic, and have better pharmacodynamic and pharmacokinetics activity [55].

• In an effort to categorise novel quinolones in the antibacterial activity against Gram-positive bacteria and their structural characteristicsBased on an understanding of their chemical structure and how this affects target selectivity, avoidance of efflux, and prevention of the formation of quinolone-resistant mutants in Gram-positive bacteria, recent and potential future advancements may be understood [56].

III. CONCLUSION

- As ciprofloxacin became the therapy of choice for many doctors seeking to treat Gram(-) infections and as the threat of multiresistant pneumococci increased demand for new medications, quinolones were launched with great fanfare in the mid-1980s.
- The effectiveness of this class will last well into the twenty-first century as long as these drugs are used to treat the right kinds of patients and are not viewed as a panacea by prescribers.

However, if they are ignored without showing any care, their day would end too soon.

• Since bacteria are constantly more intelligent than humans, conserving antibiotics as effective treatments rather than historical discoveries calls for both theoretical and highly practical approaches.

REFERENCES

- 1) K. Coleman, Drug Discov. Today, Therapeutic Strategies 2004, 1, 455–460.
- S. Emami, A. Shafiee, A. Foroumadi, Mini Rev. Med. Chem. 2006, 6, 375–386.
- 3) J. Fung-Tomc, B. Minassian, B. Kolek, T. Washo, et al., J Antimicrob. Chemother. 2000, 45, 437–446.
- F. Garcia-Garrote, E. Cercenado, J. Martin Pedroviejo, O. Cuevas, E. Bouza, J. Antimicrob. Chemother. 2001, 47, 681–684.

- J. H. Kim, J. A. Kang, Y. G. Kim, J. W. Kim, et al., Antimicrob. Agents Chemother. 1997, 41, 2209– 2213.
- Ball, P.; Mandell, L.; Niki, Y.; Tilloston, G. Drug Safety 1999, 21, 407.
- 7) Ball, P. J. Antimicrob. Chemother. 2000, 45,557.
- Be Sarro, A.; De Sarro, G. Curr. Med. Chem 2001, 8, 371.
- 9. Anderson, V. E.; Osheroff, N. Curr. Pharm. Des. 2001, 7, 339.
- Hong, C. Y.; Kim, Y. K.; Chang, J. H.; Kim, S. H.; Choi, H.; Nam, D. H.; Kim, Y. Z.; Kwak, J. H. Novel Fluoroquinolone Antibacterial Agents Containing Oxime-Substituted (Aminomethyl) pyrrolidines: Synthesis and Antibacterial Activity of 7-(4-(Aminomethyl)- 3(methoxyimino)pyrrolidin-1-yl)-1cyclopropyl-6-fluoro-4-oxo-1,4 dihydro [1,8]naphthyridine-3-carboxylic Acid (LB20304). J. Med. Chem. 1997, 40, 3584-3593.
- Radl, S.; Kovarova, L. Synthesis and Antibacterial Activity of New 7-Substituted Fluoroquinolones. Collect. Czech. Chem. Commun. 1991, 56, 2406-2412.
- Cooper, C. S.; Klock, P. L.; Chu, D. T. W.; Hardy, D. J.; Swanson, R. N.; Plattner, J. J. Preparation and in Vitro and in Vivo Evaluation of Quinolones with Selective Activity against Gram-Positive Organisms. J. Med. Chem. 1992, 35, 13392-1398.
- 13) Foroumadi, A.; Emami, S.; Davood, A.; Moshafi, M. H.; Sharifian, A.; Tabatabaie, M.; TarhimiFarimani, H.; Sepehri, G.; Shafiee, A. Synthesis and in vitro antibacterial activities of N-substituted piperazinylquuinolones. Pharm. Sci. 1997, 3, 559-563.
- 14) Hong, C. Y.; Kim, Y. K.; Lee, Y. H.; Kwak, J. H. MethyloximesubstitutedAminopyrrolidine: A New Surrogate for 7-Basic Group of Quinolone. Bioorg. Med. Chem. Lett. 1998, 8, 221-226.
- 15) Foroumadi, A.; Emmami, S.; Haghighat, P.; Moshafi, M. H.Synthesis and In-vitro Antibacterial Activity of New N-Substituted Piperazinyl Quinolones. Pharm. Pharmcol. Commun. 1999, 5, 591-594.
- 16) R.E. Polk, Am. J. Med. 87 (1989) S76. Hooper, D. C. Lancet Infect. Dis. 2002, 2, 530.
- 17) Beermann, D.; Kuhlmann, J.; Dalhoff, A.; Zeiler, H. J.
- 18) Quinolone Antibacterials; Springer: New York, 1998.
- 19) Atarashi, S.; Imamura, M.; Kimura, Y.; Yoshida, A.; Hayakawa, I.; Fluorocyclopropyl Quinolones. 1. Synthesis and Structure-Activity Relationships of 1-(2 Fluorocyclopropyl) -3 pyridone carboxylic Acid

Antibacterial Agents. J. Med. Chem. 1993, 36, 3444-3449.

- 20) J.-Y. Fan, D. Sun, H. Yu, S.M. Kerwin, L.H. Hurley, J. Med. Chem.38 (1995) 408.
- S. Dessus-Babus, C.M. Bebear, A. Charron, C. Bebear, B. De Barbeyrac, Antimicr. Agent Chemother. 42 (1998) 2474.
- 22) S. Ameyama, Y. Shinmura, M. Takahata, Antimicr. Agent Chemother.47 (2003) 2327.
- 23) D.E. King, R. Malone, S.H. Lilley, Am. Fam. Phys. 61 (2000) 2741.
- 24) E.K. Efthimiadou, M. Katsarou, Y. Sanakis, C.P. Raptopoulou, A. Karaliota, N. Katsaros, G. Psomas, J. Inorg. Biochem. 100 (2006) 1378.
- 25) E.K. Efthimiadou, Y. Sanakis, C.P. Raptopoulou, A. Karaliota, N. Katsaros, G. Psomas, Bioorg. Med. Chem. Lett. 16 (2006) 3864.
- 26) G. Psomas, A. Tarushi, E.K. Efthimiadou, Y. Sanakis, C.P. Raptopoulou, N. Katsaros, J. Inorg. Biochem. 100 (2006) 1764.
- 27) E.K. Efthimiadou, H. Thomadaki, Y. Sanakis, C.P. Raptopoulou, N. Katsaros, A. Scorilas, A. Karaliota, G. Psomas, J. Inorg. Biochem. 101 (2007) 64.
- 28) E.K. Efthimiadou, G. Psomas, Y. Sanakis, N. Katsaros, A. Karaliota, J. Inorg. Biochem. 101 (2007) 525.
- 29) E.K. Efthimiadou, Y. Sanakis, N. Katsaros, A. Karaliota, G. Psomas, Polyhedron 26 (2007) 1148.
- 30) E.K. Efthimiadou, N. Katsaros, A. Karaliota, G. Psomas, Bioorg. Med. Chem. Lett. 17 (2007) 1238.
 A. Tarushi, P. Christofis, G. Psomas, Polyhedron 26 (2007) 3963.
- Tarushi, E.K. Efthimiadou, P. Christofis, G. Psomas, Inorg. Chim. Acta 360 (2007) 3978.
- 32) E.K. Efthimiadou, N. Katsaros, A. Karaliota, G. Psomas, Inorg. Chim. Acta 360 (2007) 4093.
- 33) G. Psomas, A. Tarushi, E.K. Efthimiadou, Polyhedron 27 (2008) 133.
- 34) E.K. Efthimiadou, A. Karaliota, G. Psomas, Polyhedron 27 (2008) 349.
- 35) M.E. Katsarou, E.K. Efthimiadou, G. Psomas, A. Karaliota, D. Vourloumis, J. Med. Chem. 51 (2008) 470.
- 36) E.K. Efthimiadou, M.E. Katsarou, A. Karaliota, G. Psomas, J.Inorg. Biochem. 102 (2008) 910.
- 37) European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID). Determination of minimum inhibitory concentrations (MICs) of antibacterial agents by agar

dilution. Clinical Microbiology and Infection, Eucast Definitive Document E. Def 3.1, 2000, 6, 509–515.

- 38) Baron, E. J.; Finegold, S. M. Bailey and Scott_s Diagnostic Microbiology, 8th ed.; C.V. Mosby: St. Louis, 1990; pp184–188.
- 39) A.D. Russell, in: S.S. Block (Ed.), Disinfection, Sterilization and Preservation, 4th ed., Lea and Febinger, Philadelphia, 1991, p. 27.
- 40) H.W. Rossmore, in: S.S. Block (Ed.), Disinfection, Sterilization and Preservation, 4th ed., Lea and Febinger, Philadelphia, 1991, p. 290.
- 41) C. Dendrinou-Samara, G. Psomas, C.P. Raptopoulou, D.P. Kessissoglou, J. Inorg. Biochem. 83 (2001) 7.
- 42) G. Psomas, C. Dendrinou-Samara, P. Philippakopoulos, V. Tangoulis, C.P. Raptopoulou, E. Samaras, D.P. Kessissoglou, Inorg. Chim. Acta 272 (1998) 24.
- 43) E.K. Efthimiadou, M. Katsarou, Y. Sanakis, C.P. Raptopoulou, A. Karaliota, N. Katsaros, G. Psomas, J. Inorg. Biochem. 100 (2006)1378
- 44) E.K. Efthimiadou, M.E. Katsarou, A. Karaliota, G. Psomas, J. Inorg. Biochem. 102 (2008) 910.
- 45) Wise, R.; Andrews, J. M.; Edwards, L. J. In Vitro AntibacterialActivity of Bay 09867, a New Quinolone Derivative, Compared with Those of Other Antimicrobial Agents. Antimicrob.AgentsChemother. 1983, 23, 559-564. (b) Grohe, K.; Heitzer,H. Cycloaracylation of Enamines, I. Synthesis of 4-Quinolone-3-carboxylic Acids. Liebigs Ann. Chem. 1987, 29-37.
- 46) Atarashi, S.; Yokohama, S.; Yamamzaki, K.; Sakano, K.;Imamura, M.; Hayakawa, I. Synthesis and Antibacterial Activityof Optically Active Ofloxacin and Its FluoromethylDerivative.Chem. Pharm. Bull. 1987, 35, 1896-1902. (b) Une, T.; Fujimoto,T.; Sato, K.; Osada, Y. Synthesis and Antibacterial Activity ofOptically Active Ofloxacin. Antimicrob. Agents Chemother. 1988,32, 559-564.
- 47) Asahina, Y.; Ishizaki, T.; Suzue, S. Recent advances instructure activity relationships in new quinolones. In FluorinatedQuinolonessNew Quinolones Antimicrobials; Mitsuhasi,S., Junker, E., Eds.; Progress in Drug Research, Vol. 38;Birkhauser Verlag: Basel, Switzerland, 1992; pp 57-106. (b)Domagara, J. M. Structure-activity and structure-side-effectrelationships for the quinolone antibacterials. J. Antimicrob.Chemother. 1994, 33, 685-706.
- 48) Atarashi, S.; Imamura, M.; Kimura, Y.; Yoshida, A.; Hayakawa,I.; Fluorocyclopropyl Quinolones. 1.

Synthesis and Structure-Activity Relationships of 1-(2-Fluorocyclopropyl)-3-pyridonecarboxylic Acid Antibacterial Agents. J. Med. Chem. 1993, 36,3444-3448.

- 49) European Committee for Antimicrobial Susceptibility Testing (EUCAST)of the European Society of Clinical Microbiology and InfectiousDiseases (ESCMID). "Clinical Microbiology and Infection," EucastDefinitive Document E. Def 3.1, 2000, 6, pp. 509—515.
- 50) Japan Society of Chemotherapy. Chemotherapy 1981, 29, 76.
- 51) Niedt, G. W.; Schinella, R. A. Arch. Pathol. Lab. Med. 1985,109, 727.
- 52) Graul, A.; Rabasseda, X.; Castaner, J. Drugs Fut. 1999,24, 1324.
- 53) Elmore, S. W.; Cooper, C. S.; Schultz, C. C.; Hutchinson, D. K.; Donner, P. L.; Green, B. E.; Anderson, D. D.; Xie, Q. WO 01/ 32655 A2, 2001, Chem. Abstr. 2001, 134, 34037.
- 54) . Lohray, B. B.; Lohray, V. B.; Srivastava, B. K.; Kapadnis,P.; Pandya, P. Bioorg. Med. Chem. 2004, 12, 4557.National Committee for Clinical Laboratory Standards.Methods for Dilution Antimicrobial Susceptibility Tests forBacteria That Grow Aerobically, 4th ed. (ApprovedStandard); NCCLS Document M7-A4; NCCLS: Wayne,Pa. 1997.
- 55) Yamamoto, T.; Miura, T.; Isomura, Y.; Tadauchi, K.;Iida, H.; Ouchi, S.; Katano, K. WO 97/ 36887, 1997,Chem. Abstr. 1997, 127, 307400; (b) Halczenko, W.;Stump, C. A. WO 99/ 27929, 1999, Chem. Abstr. 1999,131, 19011; (c) Koike, H.; Asai, F.; Sugidachi, A.; Kimura,T.; Inoue, T.; Nishino, S.; Tsuzuki, Y. CA 2077695, 1994,Chem. Abstr. 1994, 120, 8583.