

# Antimicrobial and phytochemical evaluation of ascidian *Phallusia nigra* from Vizhinjam Bay, Southwest coast of India

Victor Shanthini Deepa<sup>1</sup>, Gnanakkan Ananthan<sup>2</sup>, Rajaram Murugan<sup>3</sup>, Anandakumar Arunkumar<sup>4</sup>

Centre of Advanced Study in Marine Biology, Faculty of Marine Sciences  
Annamalai University, Parangipettai-608502, Tamil Nadu, India.

**Abstract-** The antimicrobial activity of methanol, ethanol, chloroform, aqueous and ethyl acetate were carried out by using disc diffusion method against various pathogenic organisms. The crude ethanol extract was more active exhibiting a broad spectrum of antibacterial activity. Maximum Inhibition zone (16mm) was observed against *Staphylococcus aureus* in 40 µg/ml concentration of ethanol extract. The minimum inhibition zone (7mm) was observed in chloroform extract of streptococcus mutans against 40 µg/ml concentration. Among the antifungal activity maximum Inhibition zone 17mm was observed in *Aspergillus flavus* of ethanol extract and Minimum Inhibition (7mm) was observed in *Rhizopus* species of methanol and chloroform extract of 40 µg/ml concentration. The preliminary phytochemical screening of various extracts was carried out. The extractive value was Maximum in protein (ie) 120 µg/ml and 65 µg/ml in flavonoid of ethanol extract and minimum 4 µg/ml in Tannin of ethanol extract. Alkaloids, steroids, flavonoids, tannin, saponins, carbohydrates, amino acids, proteins, phenols and lipids are present. FTIR results showed that mostly phenolic compound carboxylic acids and alkane group present in ascidian *Phallusia nigra*.

**Keywords-** *Phallusia nigra*, Antimicrobial, Antifungal, Phytochemical screening, FTIR.

## I. INTRODUCTION

The ocean is considered to be an untapped source for many things including potential drugs. A large proportion of natural compounds have been extracted from marine invertebrates especially sponges, ascidians, bryozoans and molluscs and some of them are currently used in clinical trials (Proksch et al., 2002). Ascidians are marine invertebrates which ranks second with promising the source of drugs (Azumi et al., 1990). Most of the ascidians are utilized as food in various countries and they are known to produce bioactive metabolites which prevent bio-fouling and this can be considered as a kind of autogenic protection (Barguest and

Bed ford, 1978). Tunicates have been reported as rich sources of biologically active compounds and ranked third for their overall activities next to sponges and bryozoans (Davis and Bremner, 1999). These compounds are mainly comprised of various derivatives of alkaloids and peptides. There are few examples of marine derived compounds which have successfully reached the market as therapeutic drugs. A large group of low molecular weight natural compounds that exhibit antimicrobial activity has been isolated from plants and animals during the past two decades. Polyphenols and other phytochemical have been shown to have antioxidant activities, it has been suggested that consumption of polyphenol-rich foods is associated with reduced risk of cardiovascular diseases, stroke and certain types of cancer (Borros et al., 2007 and Jegadish et al., 2003).

However the phytochemical analysis using these extracts of *phallusia nigra* has not been carried out so far.

## II. MATERIALS AND METHODS

### Collection and preparation of sample

The ascidian, *Phallusia nigra* (Chordate: Ascidiacea) was collected during the low tide of the intertidal area at Vizhinjam south coast of India during April 2017. The collected samples were rinsed with sterile sea water to remove associated debris and salt. The samples were weighed (10g) and preserved separately in methanol and ethanol (1:2) and brought to the laboratory

### Preparation of the extract

The test samples were carefully removed washed with sterile seawater dried under shade and homogenized to get a coarse powder. The coarse powder was stored in an air tight container and used for further investigation. 100g of the powdered animal material was extracted with methanol, ethanol, chloroform, ethyl acetate and aqueous using soxhlet apparatus. The extract was cooled to room temperature,

evaporated in a rotary evaporator under reduced pressure and a brown sticky residue was obtained (15g).

### Antibacterial assay

The bioassay was carried out using the agar disc diffusion method (Bauretal., 1966). Muller Hinton agar plates are prepared by pouring 15ml of medium and allowed to solidify. The petriplates are swabbed with 24 hour old culture of the selected bacterial strains. The sterile paper discs were loaded with different solvents concentration and allowed to dry thoroughly. Then the discs were placed over the plates and incubated from 24hr at 37°C.

### Antifungal assay

The bioassay was carried out using Ratiram et al., (2015). Zobell dextrose agar medium were prepared and the compound was allowed to diffuse for 5 minutes and the plates were kept for incubation at 37°C for 24 hours. At the end of incubation inhibition zones were examined.

### Phytochemical analysis

#### Qualitative analysis and Quantitative analysis

The following phytochemicals were qualitatively and quantitative determined in the ascidian *Phallusia nigra* among the 5 different extracts.

Carbohydrates estimated by Fehling's Test (Kokate,1994),Glycosides (Ansari 2006) by Keller – Killiani test), Steroids (IP,1996) by Salkowski Test, Alkaloids (Ansari 2006) byMayers Test, Flavanoids (kokate, 1994) by Shinoda Test, Tannin (Mukherjee, 2002) by Lead acetate test, Saponin (Ansari, 2006) byFoam test, Protein (Ansari, 2006) by Biuret test, Phenol by Ferric chloride test (Mutherjee, 2002),Amino acid by Ninhydrin (Ansari 2006). Are the test used for qualitative andquantitative determination.

### FTIR

Methanol, ethanol, chloroform, ethyl acetate and aqueous extract of *Phallusia nigra* were recorded by Fourier-Transformed Infrared Spectra (Shimadzu,UK). FTIR was used to identify the chemical structure in a wide range of compounds. Infrared spectroscopy was a useful analytical tool for detection of functional groups in organic compounds. IR spectra were recorded in the 400-4000 cm<sup>-1</sup> with a resolution of km<sup>-1</sup>. The room was kept at a controlled ambient temperature (25°C) and relative humidity 30% (Sayed et al., 2005).

## III. RESULTS AND DISCUSSION

The results of antibacterial activity and antifungal activity of the crude methanol, ethanol, ethyl acetate, chloroform and aqueous extract of *Phallusia nigra* against 3gram positive and 3gram negative bacteria (Table 1).

Table 1 Antibacterial and Antifungal activity of aqueous extract from ascidian *Phallusia nigra*

Sample Name	ANTIBACTERIAL						ANTIFUNGAL		
	<i>Staphylococcus</i>	<i>Streptococcus</i>	<i>Bacillus subtilis</i>	<i>Klebsiella</i>	<i>Proteus vulgaris</i>	<i>Escherichia coli</i>	<i>Aspergillus niger</i>	<i>Aspergillus flavus</i>	<i>Rhizopus stolonifer</i>
Ethanol	16	13	11	11	8	9	13	17	11
Methanol	10	10	10	11	9	12	14	14	7
Ethylacetate	9	9	NZ	NZ	8	NZ	NZ	9	NZ
Chloroform	8	7	10	NZ	NZ	NZ	NZ	13	7
Aqueous	NZ	NZ	NZ	NZ	NZ	8	8	NZ	NZ
Positive Control (streptomycin)	25	26	18	26	21	26	26	21	17

Ethanol extract of 40ug/ml concentration produced maximum inhibition zone of 16mm against *Staphylococcus aureus* and minimum of 7mm in *Streptococcus mutans*. Corresponding zones of methanol extract produced 12mm and 11mm against *E. coli* and *Proteus vulgaris* (Table 1). All the 5 extracts of some concentration of different organisms showing minimum activity against ethylacetate and chloroform against *Staphylococcus aureus*, *Streptococcus mutans*, *E. coli*, *B. subtilis*, *K. pneumoniae*, *P. vulgaris*. This observation is consistent with the findings of Ananthan et al., (2011) who reported that both methanol and ethyl acetate extract of *P. nigra* showed a broad spectrum of antibacterial activity against tested gram negative pathogens. Antibacterial activity of ascidians extract increased with increasing concentrations. Antibacterial activity of some tunicates has been previously reported from various authors of Thompson *et al.* 1983, Mohamed Hussain and Ananthan, 2009, Sivaperumal et al., 2010, Ananthan et al, 2011, Mohamed Hussain et al, 2004. The results of the present study showed that highly potent antibacterial substance is found in

*Phallusia nigra*. Antibacterial activity of crude extract of ascidian showed inhibiting activity against almost all the 6 human pathogen strain. The range of inhibiting distance of bacteria varied from 10-11mm. (Table1)

Antifungal activity of 5 different extracts of ascidians shows that maximum Inhibition zone of 17mm noticed in *Aspergillus niger* at ethanol extract and minimum Inhibition zone of 7mm noticed in chloroform and methanol extract of *Rhizopus stolonifer* (Table 1).

The present findings of Neda et al., 2014, antifungal activity of *Phallusia nigra* of Persian Gulf shows that maximum inhibition zone of 25mm observed in *Aspergillus niger* when the concentration increased from 5 to 20 $\mu$ l.

Table 2 Preliminary phytochemical screening of the various extracts of *Phallusia nigra*

S. No	Test	Ethanol	Methanol	Ethyl acetate	Chloroform	Aqueous
1.	Carbohydrate	-	-	-	+	-
2.	Protein	+	+	-	-	-
3.	Amino acid	+	-	-	-	-
4.	Alkaloid	+	+	-	-	-
5.	Flavonoid	+	+	-	-	-
6.	Glycoside	-	-	+	+	-
7.	Steroid	-	-	-	-	-
8.	Saponin	+	-	-	-	-
9.	Tannin	+	-	-	-	+
10.	Phenol	-	-	-	+	-

The phytochemical constituents like alkaloids, flavonoids, steroids, tannins, saponin, phenols, proteins, amino acids, carbohydrate, Glycosides were tested using 5 different extracts like ethanol, methanol, chloroform, aqueous and ethyl acetate. The results are presented in table 2. The ethanol and methanol showed the presence of alkaloid, and carbohydrate. The methanol extract showed the presence of alkaloids, flavonoids, proteins and amino acids. Aqueous extract showed the presence of phenols, carbohydrate and tannin

The knowledge of phytochemical screening is desirable, not only for the discovery of therapeutic agents but also because such information may be of value disclosing new sources of such economic materials, like tannins, oils, gums, precursor for the synthesis of complex chemical substances etc. Major phytochemical constituents like alkaloids, steroids, flavonoids, saponins are present in almost all extraction. Phenolic compounds saponins and flavonoids may be linked or suggested to be involved in antibacterial, antiviral and anti diarrheal activity as suggested by Majaw and Moirangham in plants. Investigations on the mode of action by Enzo in plants indicate that flavonoid increase colonic water, electrolytic re absorption and other chemicals act by inhaling intestinal

mobility while some components have been show to inhibit particular enter pathogens. Alkaloids are reported to have cardiovascular effects by Juge et al. (2001). As ascidians are sedentary animals the same role may be suggested in these animals as defense mechanism.

Table 3 Quantitative estimation of phytochemicals

Extract	Glycoside ug/ml	Flavonoid ug/ml	Saponin ug/ml	Protein ug/ml	Tannin ug/ml	Phenol ug/ml	Carbohydrate ug/ml	Amino Acid ug/ml
Methanol	-	25	-	55	-	-	-	-
Ethanol	-	65	39	120	4	-	-	46
Chloroform	10	-	-	-	-	39	20	-
Ethyl acetate	13	-	-	-	-	-	-	-
Aqueous	-	-	-	-	5	-	-	-

Phytochemicals like glycoside, flavonoid, saponin, protein, phenol, carbohydrate and Amino acids are present in ascidian *Phallusia nigra*. Maximum production is protein 120 $\mu$ g/ml in ethanol extract. Second maximum production is flavonoid 65 $\mu$ g/ml of ethanol extract and minimum production of Tannin 4 $\mu$ g/ml of ethanol extract only. (Table 3)

Phytochemicals play a vital role in the medicinal properties. Consumption of spices has been implicates in the prevention of many chronic diseases such as cardiovascular diseases, cancer and inflammation. (Panpati et al, 2013). Phenolic are good antioxidants and exhibit a wide range of pharmacological properties such as anticancer, anti-inflammatory and anti-diabetic effects (Hamazah et al., 2013). Flavonoids are group of more than 4000 polyphenol compounds that occur naturally in food of plant origin, possessing a number of beneficial effects to human health (Menaga et al., 2012). They are potent water soluble, super antioxidants and free radical scavengers that prevent oxidative damage, and have anticancer, antiinflammatory, anti allergic, antiviral, vasodilating properties and inhibition of platelet aggregation. Also it is known that saponins inhabitant efflux by blockage of the influx of concentration in the cells activating a Na<sup>+</sup>-Ca<sup>2+</sup> antiporter in cardiac muscles. The increase in Ca<sup>2+</sup> influx through this anti porter strengthens the contraction of cardiac muscles (Ergwim et al., 2011; Schneider et al., 2004)

Tannins may elicit antibacterial activities via cell membrane lysis, inhibition of protein synthesis, proteolytic enzymes and microbial adhesions. (Dulgar et al., 2002).Tannins is used to treat nonspecific diarrhoea and inflammation of the mouth).

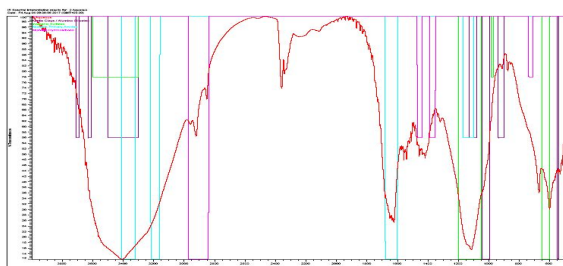


Fig. 1 FTIR Spectrum of the aqueous extract of *Phallusia nigra*

Table 4 Peak values of aqueous extract of *Phallusia nigra*

Frequency ranges	Intensities	Assignment and remarks	Group or Functional Class
3404	Medium	O-H stretch	Phenols, alcohol
2359.47	Strong	O-H stretch	Carboxylic acids

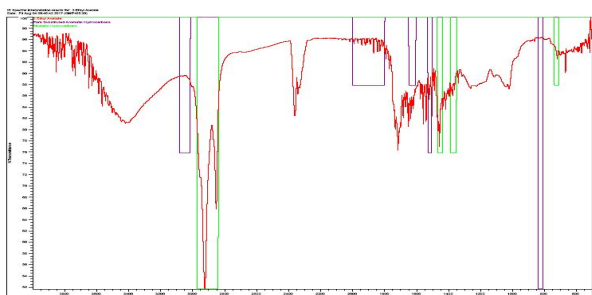


Fig. 2 FTIR Spectrum of Ethyl acetate extract of *Phallusia nigra*

Table 5 Peak values of FTIR Ethyl acetate extract of *Phallusia nigra*

Frequency ranges	Intensities	Assignment and remarks	Group or Functional Class
2924.15	Strong	H-C-H Asymmetric stretch	Alkane
2362.20	Strong	Hydrogen bonded O-H stretch	Carboxylic acids

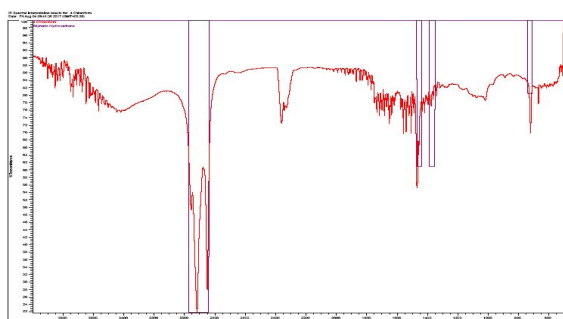


Fig.3 FTIR Spectrum of chloroform extract of *Phallusia nigra*

Table 6 Peak values of FTIR chloroform extract of *Phallusia nigra*

Frequency ranges	Intensities	Assignment and remarks	Group or Functional Class
2917.17	Strong	H-C-H- Asymmetry and Symmetry	Alkane
2819.76	Strong	H-C-H- Asymmetry and Symmetry	Alkane
2360.39	Strong	Hydrogen bonded O-H stretch	Carboxylic acids

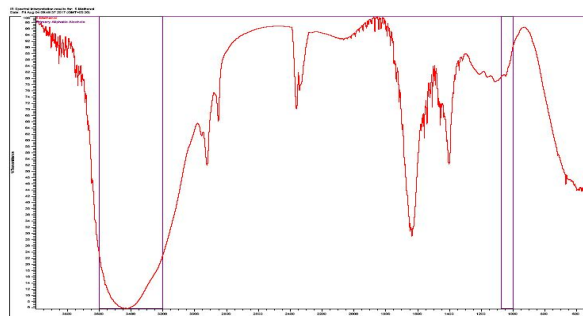


Fig. 4 FTIR Spectrum of Methanol extract of *Phallusia nigra*

Table 7 Peak values of the FTIR Methanol extract of *Phallusia nigra*

Frequency ranges	Intensities	Assignment and remarks	Group or Functional Class
3444.15	Strong	N-H stretch	Amines
2359.72	Strong	Hydrogen bonded O-H stretch	Carboxylic acids
1716.80	Medium	C=O stretch	Ketone
1404.24	Strong	H-C-H Bond	Alkane

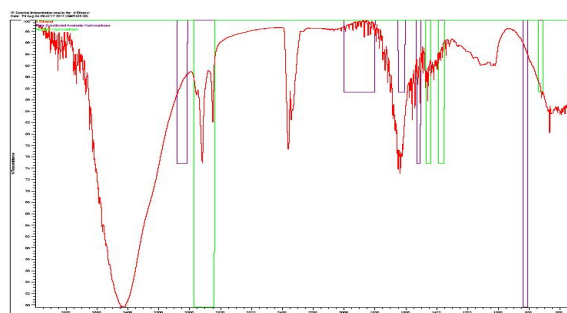


Fig. 5 FTIR Spectrum of Ethanol extract of *Phallusia nigra*

Table 8 Peak values of the FTIR Ethanol extract of *Phallusia nigra*

Frequency ranges	Intensities	Assignment and remarks	Group or Functional Class
3445.11	Strong	N-H stretch	Amines
2918.11	Strong	C-C-H Asymmetric stretch	Benzene Aromatic rings
2359.72	Strong	Hydrogen bonded O-H Stretch	Carboxylic acids
1500.4	Medium	N=O stretch	Nitro group

Aqueous extract of *Phallusia nigra* was analysed by FTIR (Fig. 1). It has phenol and alcohol with OH stretch (3404) carboxylic acids OH (2359.47) (Table 4). *P.nigra* ethylacetate extract analysed by FTIR (fig2) It has the compound Alkane H-C-H stretch (2924.15) carboxylic acids with O-H stretch (Table-5) *P.nigra* chloroform extract were analysed by FTIR (Fig. 3) it has the alkane group H-C-H Asymmetry and symmetry (2917.17), (2819.76) and carboxylic acids –O-H stretch (2360.39) (Table 6). *P.nigra* methanol extract was analysed by FTIR (Fig. 4) it has the amines N-H stretch (3444.15) carboxylic acids hydrogen bonded O-H stretch (2359.72) ketone with C=O stretch (1716.80) and alkane H-C-H bond (1404.24) (Table7). *P.nigra* of ethanol extract was analysed by FTIR (Fig. 5) it has the benzene Aromatic rings with C-C-H stretch (2918.11) carboxylic acids O-H stretch (2359.72) and nitro group with N=O stretch (1500.4) (Table8).

However crude extracts of ascidians *Phallusia nigra* with many compounds and their active portion may be very low. FTIR major peaks showed that it have phenol, methane, ketone and carboxylic acid group as a major groups in the ascidians. Phenolic compounds exhibited good antioxidant and antimicrobial activities.

#### REFERENCES

- [1] Majaw.S., Moirangthem,J.,2009 Qualitative and Quantitative analysis of (clerodendron colebrokianum walp) leaves and zingiber cassumunar Roxb. Rhizomes Ethnobotanical leaflets, 2009 13 :578-589.
- [2] Enzo A.P., Traditional plants and herbal remedies used in the treatment of diarrhoeal disease. Mode of action, quality, efficacy and safety considerations Modern Phytomedicine turning medicinal plants into drugs WILEY-VCH verlag Gmbhand co K Ga A weinheim 2007, pp: 248-260.
- [3] Juge, M., Grimaud,N.,Biard,J.E.,sauviat,M.P.,Nabil, M.,Verbist,J.F.,and Petit, J.Y.,Cardiovascular effects of lepadiformine, an alkaloid isolated from the ascidian *Clardina lepadiformis* and *C.moluccensis* Toxicon, 2001,39:1231-1237.
- [4] Udu-Ibiam, O.E., Ogbu, O., Ibiam, U.A.,Nnachi U., Agah,M.V., aigbu, U.K.,Chukwu, C.O., Agumah, O.S.,Ogbu, N.B., phytochemical and Antioxidant analysis of selected Edible Mushrooms , Gingerand Garlic from Ebonyl state, Nigeria. Journal of pharmacy and Biological sciences2014.PP.86-91.
- [5] Hamzah, R.U.,Egwim ,E.C.,Kabiru, A.Y., Muazu, M.B., Phytochemical and invitro antioxidant properties of the methanolic extract of fruits of *Blighia sapida*, *vitellaria paradoxa* and *vitex doniana* oxid antioxiid Med sci 2013 2 (3): 215-221.
- [6] Nagavani, Y., Madhavi,R.D., Bhaskar, R.P., Koteswara and Raghava.R.T., free radical scavenging activity and qualitative analysis of polyphenols by RP-HPLC in the flowers of (*couroupita guianensis* abul Electron) Environ Agric food chem, 9(9) 2010, 1471-1481.
- [7] Hamzah, R.U., Jigamu, A.A., Makun,H.M., and Egwin E.C., Phytochemical screening and antioxidant activity of methanolic extract of selected wild edible Nigerian Mushrooms, Asian specific of tropical disease, 4CD,2014,5153-5157.
- [8] Menaga, D., Mahalingam, P.U., Rajakumar S., and Ayyasamy,P.M.,Evaluation of phytochemical characteristics and Antimicrobial activity of *pleurotus Flarida* Mushrooms Asian Journal of Pharmaceutical and clinical research , 5(4) : 2012, 102-106.
- [9] Idns, S., Ndukwe, G.I., and Gimba, C.F., Preliminary Phytochemical screening and antimicrobial activity of seed extracts of *Persea americana* (AVOcado pear) Bayero journal of pure and applied sciences 2(1) 2009,(173-176).
- [10]Alai, Z., Arapour,M., and Mohensi M., Inhibitory effect of ginger extract on *C.albicans*, American Journal of applied science J, 2009,10-12.
- [11]Egwim, E.C., Ellen,R.C., and Egwach, R.U., Proximate composition , phytochemical screening and antioxidant activity of ten selected edible mushroom, American journal of food and Nutrition 1 (2), 2011, 89-94.
- [12]Schneiderand, G., Wolfing,J., Synthetic Cardenolides and related compounds, current organic compounds8, 2004.
- [13]Schulte, R.E.,and Raffaur, R., The healing forest medicinal and toxic plants of the north west Amazon Portland RF pioscomds press, 1990, 32-36.
- [14]Dulger, B., Ergul, C.C., and Guun, F., Antimicrobial activity of the macrofungus *lepistanuda fitoterpia*, 73,2002, 695-697.
- [15]Westernardp, H., Effect of tanninsin animal nutrition Dtsch tierarzt wochenshm, 113(7), 2006, 264-269.
- [16]Ananels T.P., and Edwards, J.K.P., Antimicrobial activity in the tissue extract of the five species of cowries *cyprea* spp.Mollusca, Gastrpoda and an ascidian *Didemnum psammatodes* Tunicata *Didemnidae*, Indian Journal of marine science 31(3) 2002, 239-242.
- [17]Bao, H.N., Ushio,H., and Oshima Antioxidant activity and anti discoloration efficiency of ergothionine in mushroom (*Flammulina veluptes*) extract added to beef and fish meats Journal of agricultural and food chemistry , 56(21) 2008, 10032-10040.
- [18]Hamzah, R.U., Jigamu,A., Makun, H.M., and Eguim,E.C., Phytochemical screening and antioxidant activity of methnolic extract of selected wild edible

- nigirian mushrooms, Asian pacific journal of Tropical disease, 4CD,2014, \$153-\$157.
- [19] Azumik Yoshimizu, M., Suzukis, Ezura and Yokosawa, H., Inhibitory effects on halocyanin an antimicrobial substance from ascidian hemocytes, on the growth of fish viruses and marine bacteria cell Mol Life Sci – 1990, 46(10) 1066-1068.
- [20] Proksch, P., Edrada, A., Ebel, R., Drugs from the Sea current status and Microbiological implications, Appl. Micro. Biotech 2002, 59; 125:134.
- [21] Bergquist Pr. Belford JJ., The incidence of antibacterial activity Marine demospongiae, Systematic and geographic considerations Mar Biology 1978, 46(3):215 – 221.
- [22] Davis A.R., Bremner, J.B., Potential Antifouling Natural Products from Ascidian A Review In M F, Sarojini and R. Nagabhusharanam (Eds) Bioactive Compounds from Marine Organisms Thompson oxford and IBH Publishing co Pvt Ltd, New Delhi. 1999, PP. 259-310.
- [23] Mohamed Hussain, S., Ananthan, G., Antimicrobial activity of the crude extracts of compound ascidians, *Didemnum clinides* and *Trididemnum (Savignii)*. Journal of Pharmacy Research 2011 4(7):2032 – 2033.
- [24] Sivaperumal, P., Ananthan, G., Mohamed Hussain, S., Exploration of antibacterial effects on the crude extracts of marine ascidian *Aplidium multiplicatum* against Clinical isolates International Journal of Medicine and Medical Sciences 2010, 2(12) 382 – 386.
- [25] Meenakshi, V.K., Occurrence of a new ascidian Species – *Distaplia nathensis* sp. nov and two species *Eusynstyela tin* (Van Name 1902) *Phallusia nigra* (Savigny, 1816) new records for Indian Waters, Indian J. Mar. sci 1998, 27 : 477.
- [26] Sayed, Ismail, J., Ahmed, O.H., Abd E Samei, Y.M., Asker. M. Studies on the Production of Sulfated Polysaccharide by locally isolated bacteria Egyptian Pharm J 2005, 4, 439 – 452.
- [27] Medical, M.H., Salama., Najat Marraik., Antimicrobial activity and Phytochemical analysis of *Polygonum articulare* L (Polygonaceae) naturally growing in Egypt Saudi I Biol Sci 2010, 17 : 57 – 63.
- [28] Kostic, D.A., Velickovic, J.M., Mitic, S.S., Mitic, M.N., Ranedlovic, S.S., Phenolic Content and antioxidant and antimicrobial activities of *Ortaegus Oxyacantha* L , Fruit extract from South east Serbia. Trop J Pharma Res 2012; II (i) : 117 – 124.