Leishmania-Host Interaction and Survival Strategies "A Review"

Dr. Heena Sachdeva, Dr. Manish Sharma

Abstract- In order to develop a successful parasitic relationship with its host, Leishmania parasites use to interact and infect a number of host cell types viz. macrophages and dendritic cells for the successful parasitic relationship with its host. During the initial stage of the infection, both neutrophils and macrophages are recruited at the site and these acts as intermediate hosts to be used by parasites to enter macrophages.

I. INTRODUCTION

The uptake of Leishmania promastigotes by host cells is a classical receptor-mediated process that initiates phagocytosis. The complement receptors (CR)1, CR3 (Mac-1) fibronectin receptor and mannose-fucose receptor on the surface of macrophages play important roles in promastigote binding [1]. The surface lipophosphoglycan (LPG), GP63 and proteoglycans on L. major promastigotes are also important determinants in the initiation of phagocytosis and intracellular survival of parasites [2,3].

Macrophages are the major effector cells responsible for destruction of the parasites. Macrophages can be activated by different signals and their activation is generally divided into classical and alternative types. Classical activation is mediated by the products of Th1 and NK cells, in particular, IFN-gamma. IFN-gamma stimulates macrophages to produce inducible nitric oxide synthase (iNOS), an enzyme which catalyzes L-arginine to generate nitric oxide [4]. NO is a toxic molecule that plays a major role in killing intracellular parasites, including Leishmania [5].

In contrast to classical activation, alternative macrophage activation is induced by Th2 cytokines such as IL-4 and IL-13 [6]. It has been found that IL-4 induced polyamine biosynthesis (via upregulation of arginase) favours L. major parasite survival in macrophages [7]. This finding reinforced the notion that Leishmania encoded arginase is a virulent factor and its expression functions to preferentially enhance alternative macrophage activation leading to parasite survival [8,9]. Several reports show a central role of dendritic cells (DC) in generating immune responses against Leishmaniasis [10]. The skin contains three DC populations that consist of epidermal langerhan cells and two migratory dermal DC subsets. Previous studies showed that epidermal

langerhan cells phagocytose L. major in vivo and migrate to draining lymph nodes for presentation to antigen specific T cells [11]. Interaction of DC with Leishmania results in IL-12p70 production [12]. Moreover, different DC subsets are differentially permissive to Leishmania parasites and thus seem to be inversely correlated with the ability of infected cells to produce IL-12p70 [13]. The production of IL-12 by DC's initiates Th1 response and protective immunity by promoting early NK cell activities (including IFN-gamma production and cytotoxicity) [14].

Leishmania may impair host macrophage signaling pathways to disrupt cellular functions. Impaired responsiveness to IFN-gamma, lipopolysaccharide (LPS), and activators of protein kinase C (PKC) have been seen in Leishmania infections [15]. Altering signal transduction through the disruption of cellular phosphorylation, either by an alteration of cellular kinases and phosphatases or by Leishmania expressing its own phosphatases that act on macrophage proteins, is used by Leishmania to enhance survival [16]. L. donovani has been shown to impair tyrosine phosphorylation and activation of JAK1, JAK2, and STAT1 in response to IFN-gamma, possibly involving the activation of the cellular protein tyrosine phosphatase SHP-1 [17,18].

Macrophages produce nitric oxide through the induction of iNOS, in response to extracellular signals, including IFN-gamma and LPS [19]. GIPLs (Glycosylinositol phospholipids) on the amastigote surface can inhibit NO production, thus, reducing leishmanicidal activity [20]. The repeating units of LPG may also protect promastigotes from toxic oxygen metabolites generated during the macrophage oxidative burst by scavenging hydroxyl radicals and superoxide anions [21]. LPG protects the parasites by attenuation of the PKC-mediated induction of the oxidative burst [22]. Gp63 has also been associated with suppression of the oxidative burst [23]. Leishmania parasites may also survive by modulating macrophage cytokine production. Both promastigotes and amastigotes have been shown to downregulate macrophage IL-12 production, which is necessary for the Th1 response [24].

Leishmania are a diverse group of intracellular pathogens that have efficiently developed adaptive measures

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to ensure their survival. Not only have they developed strategies to survive inside the sand fly vector, but the parasites have also established means to survive in the vertebrate bloodstream. Leishmania effectively use the immune response of the host to target themselves for engulfment into macrophages.Once phagocytised, they manipulate the harsh environment through the inhibition of hydrolytic enzymes, toxic metabolic products, cell signaling, cytokine production, and other events. These strategies allow Leishmania to successfully undermine the host innate and acquired immune responses and promote parasite survival [25].

REFERENCES

- M.M. Kane and D.M. Mosser "Leishmania parasites and their ploys to disrupt macrophage activation". Current Opinion in Hematology 7(1), pp. 26–31, 2000.
- [2] C. Yao, J.E. Donelson and M.E. Wilson, "The major surface protease (MSP or GP63) of Leishmania sp. biosynthesis, regulation of expression and function" Molecular and Biochemical and Parasitology, 132(1), pp. 1–16, 2003.
- [3] T. Naderer and, M. J. McConville, "The Leishmaniamacrophage interaction: a metabolic perspective" Cell Microbiology, 10(2): pp. 301–308, 2008.
- [4] F.Y. Liew, Y. Li and S. Millott, "Tumour necrosis factor (TNF-alpha) in leishmaniasis. II. TNF-alpha-induced macrophage leishmanicidal activity is mediated by nitric oxide from L-arginine" Immunology 71(4): pp.556–559, 1990.
- [5] J.L. Santos, A.A. Andrade, A.A. Dias, C.A. Bonjardim, L.F. Reis, S.M. Teixeira, and M.F. Horta, "Differential sensitivity of C57BL/6(M-1) and BALB/c (M-2) macrophages to the stimuli of IFN-gamma/LPS for the production of NO:correlation with iNOS mRNA and protein expression" J. Interferon Cytokine Res. 26(9): pp.682–688, 2006.
- [6] S. Gordon, "Alternative activation of macrophages" Nat. Rev. Immunol. 3: pp. 23–35, 2003.
- [7] P. Kropf, J.M. Fuentes, E. Fahnrich, L. Arpa, S. Herath, V. Weber, G. Soler, A. Celada, M. Modolell, and I. Muller, "Arginase and polyamine synthesis are key factors in the regulation of experimental leishmaniasis in vivo" FASEB J. 19(8): pp.1000–1002,2005.

- [8] V. Iniesta, J. Carcelen, I. Molano, P.M. Peixoto, E. Redondo, P. Parra, M. Mangas, I. Monroy, M.L. Campo, C.G. Nieto, and I. Corraliza, "Arginase I induction during Leishmania major infection mediates the development of disease" Infect. Immun. 73(9): pp. 6085–6090, 2005.
- [9] V. Iniesta, L.C. Gomez-Nieto, I. Molano, A. Mohedano, J. Carcelen, C. Miron, C. Alonso, and I. Corraliza, "Arginase I induction in macrophages, triggered by Th2type cytokines, supports the growth of intracellular Leishmania parasites" Parasite Immunol. 24(3): pp. 113– 118, 2002.
- [10] B. Leon, M. Lopez-Bravo, and C. Ardavin, "Monocyte derived dendritic cells formed at the infection site control the induction of protective T helper1 responses against Leishmania" Immunity 26(4): pp.519–531, 2007.
- [11] H. Moll, H. Fuchs, C. Blank, and M. Rollinghoff, "Langerhans cells transport Leishmania major from the infected skin to the draining lymph node for presentation to antigen-specific T cells" Eur.J.Immunol. 23(7):pp.1595–1601, 1993.
- [12] M.A. Marovich, M.A. McDowell, E.K. Thomas, and T.B. Nutman, "IL-12p70 production by Leishmania major-harboring human dendritic cells is a CD40/CD40 ligand-dependent process" J. Immunol. 164(11): pp. 5858–5865, 2000.
- [13] S. Henri, J. Curtis, H. Hochrein, D. Vremec, K. Shortman, and E. Handman, "Hierarchy of susceptibility of dendritic cell subsets to infection by Leishmania major: inverse relationship to interleukin-12 production" Infect. Immun. 70(7): pp.3874–3880, 2002.
- [14] J. Liese, U. Schleicher and C. Bogdan, "TLR9 signaling is essential for the innate NK cell response in murine cutaneous leishmaniasis" Eur. J. Immunol. 37(12): pp.3424–3434, 2007.
- [15] N.E. Reiner, "Altered cell signaling and mononuclear phago cyte deactivation during intracellular infection" Immunol.Today 15(8): pp.374–381, 1994.
- [16] M.M. Kane, and D.M. Mosser, "Leishmania parasites and their ploys to disrupt macrophage activation" Curr. Opin. Hematol. 7(1): pp.26–31, 2000.
- [17] D. Nandan, and N.E. Reiner, "Attenuation of gamma interferon- induced tyrosine phosphorylation in mononuclear phagocytes infected with Leishmania

donovani: selective inhibition of signaling through Janus kinases and Stat1" Infect. Immun. 63(11): pp.4495–4500, 1995.

- [18] M.T. Shio, K. Hassani, A. Isnard, B. Ralph, I. Contreras, M.A. Gomez, I. Abu-Dayyeh, and M. Olivier, "Host cell signaling and Leishmania mechanisms of evasion" J. Trop. Med. 2012: pp.819512, 2012.
- [19] C. Nathan, and Q.W. Xie, "Regulation of biosynthesis of nitric oxide" J. Biol. Chem. 269: 13725(19)-13728, 1994.
- [20] X.Q. Wei, I.G. Charles, A. Smith, J. Ure, G.J. Feng, F.P. Huang, D. Xu, W. Muller, S. Moncad, and F.Y. Liew, "Altered immune responses in mice lacking inducible nitric oxide synthase" Nature 375(6530) pp. 408-411, 1995.
- [21] T.B. McNeely, S.J. Turco, "Requirement of lipophosphoglycan for intracellular survival of Leishmania donovani within human monocyte". Journal of Immunology 144(7):pp. 2745–2750, 1995.
- [22] A. Descoteaux, S.J. and Turco, "Glycoconjugates in Leishmania infectivity" Biochim. Biophys. Acta 1455(2-3): pp.341–352, 1999.
- [23] A.L. Sorensen, A.S. Hey and A. Kharazmi, "Leishmania major surface protease Gp63 interferes with the function of human monocytes and neutrophils in vitro" APMIS 102 pp. 265–271, 1994.
- [24] L. Carrera, R.T. Gazzinelli, R. Badolato, S, Hieny, W. Muller, R. Kuhn, and D.L. Sacks, "Leishmania promastigotes selectively inhibit interleukin 12 induction in bone marrow-derived macrophages from susceptible and resistant mice" J. Exp. Med. 183(2): pp.515-526, 1996.
- [25] D. Liu and J.E. Uzonna, "The early interaction of Leishmania with macrophages and dendritic cells and its influence on the host immune response" Front. Cellular Infec. Microbiol. 2:83: pp.1-8, 2012.